Spatiotemporal tumor immune modulation by localized delivery of cancer therapeutics using an implantable microdevice

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INTRODUCTION

Mechanism of action studies are imperative to translating oncology therapeutics into the clinic informing testable hypotheses and potential combination partners. This often involves large and time-consuming mouse studies to test a small number of compounds to show statistically significant benefit. To reduce the time and cost for mechanism of action studies of our cytokine portfolio, we applied the implantable microdevice (IMD) NanoNail[™] developed by KiburMed to measure intra-tumoral drug responses and to differentiate the mechanism of action of multiple cytokine agents in parallel in the MC38 mouse model. With the NanoNail (IMD) device, we will compare murine surrogates of SAR445877 (mKD050) - a fusion of anti-PD1 to IL15-IL15Ra mutein - to anti-PD1 and combination of KD050 + anti-PDL1 in mouse tumors.

TECHNICAL APPROACH

C57BL/6 mice (3 groups x 11 mice per group) were subcutaneously inoculated on the flank with MC38 tumors. After sizable tumor growth, the IMD device containing 9 different therapeutic candidates was implanted. MC38-bearing mice were taken down at 24 hour, 4 days, and 7 days post IMD implantation to assess response. Pharmacodynamic response to treatments was measured by two readouts: Cyclic Immunofluorescence with 22 markers and whole transcriptome Nanostring GeoMx on regions of interest proximal and distal to tumor defined by PanCK and CD45. Among the candidate murine surrogate compounds in the device, KD050 was tested as a monotherapy, as a combination partner with anti-PDL1 and compared against anti-PD1 monotherapy.



Figure 1: Mouse MC38 tumors were grown at Charles River Labs until tumors reached an average of 350 to 500mm³. Each reservoir was filled with liquid nanodoses based on known clinical activity and implanted into each mouse tumor. Potency at nanodose level was confirmed prior to start of study. At each time point, the tumors were excised, placed in 10% formalin and transferred to 70% ethanol 24 hours post collection. The recovered IMDs were embedded in a total of 5 parrafin blocks (up to 6 IMDs per blocks). Nine thin sections were cut from each block for a total of 54 thin sections and subsequently stained for cyclic immunofluorescence and or IF markers for whole transcriptome GeoMx DSP.

CD8









Figure 3: A) Each slide representation contains four to five IMD containing tissues. Each IMD flank tissue contains tumor (white circle) and adjacent tissue. Direction of compound release depicted with arrows and markers for ROI include Dapi nuclei, PanCK for tumor, and CD45 for immune cells. Regions of interest chosen in proximal vs adjacent regions to IMD device. B) Table of tumor cores per treatment per time point. C) GeoMx whole transcriptome analytical workflow.

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RESULTS

 Combination KD050 + anti-PDL1 increases T cells (CD3, CD4, CD8) and B cells (CD20 and CD27) both by Cyclic IF and GeoMx Nanostring whole transcriptome compared to

Combination mKD050 + anti-PDL1 also increases NK cells and Cytotoxic Lymphocytes observed by Nanostring GeoMx whole transcriptome and MCP counter signatures. Most significant cellular changes were observed in proximal regions than in distal regions

Differential expression and pathway enrichment results are still ongoing.

Figure 2: Analytical Methods for Quantification of Drug Effects by Cyclic IF

Figure 2

A) Quantification of Cyclic IF Image data. IF image is split by channel and each marker is quantified within region of drug distribution and adjacent zone. B) Automated quantification pipeline counts positive cells for each marker within a given zone².



Figure 3: Experimental and Analytical Design of Spatial GeoMx DSP experiment



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REFERENCES:

- (2023) 83 (7 Supplement): 2972.
- 2. An Interactive Pipeline for Quantitative Histopathological Analysis of Spatially Defined Drug Effects in Tumors. Sebastian Ahn; Benjamin Ferland; Oliver Jonas. Journal of Pathology Informatics. 2021